Combination of *Calotropis gigantea* Radix Extract and Artemisin as an Antimalarial Agent Against *Plasmodium berghei*

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ABSTRACT

Calotropis gigantea radix is one of plant which has bioactive component as antimalarial. The purpose of this research are to know antimalarial activity from combination of Calotropis gigantea radix extract and artemisin. The research consist of extraction of Calotropis gigantea radix was done with extraction maseration method. Extraction was performed by maseration with 80% ethanol solvent. Concentrated extract was in vivo antimalarial tested to animal model. Mice were peritoneal infected with 10⁶Plasmodium berghei ANKA and divided into 5 treatment groups: negative control; positive control (artemisin of dose 4 mg/kgBW); combination of artemisin(1/4 dose of artemisin standar) and Calotropis giganteain 3 doses: 0,1 mg/KgBW; 1 mg/KgBWand 10 mg/KgBW. Data of inhibition was analyzed using SPSS program with Two Way ANOVA Test dan continued with Tukey Test.

The result shows that combination of *Calotropis gigantea* radix extract and artemisin with dose 10 mg/KgBB has higher antimalarial activity than control positive (artemisin). The value of parasite inhibition is 55,2% for dose 0.1 mg/KgBW; 72,8% for dose 1 mg/KgBW; 87,3% for dose 10 mg/KgBW and 56,06 mg/KgBW for control positive (artemisin). The phytochemical compounds in 80% ethanol solvent extract are tannins and steroids.

Keywords: Calotropis gigantea, antimalaria, combination, artemisin, Plasmodium berghei

INTRODUCTION

Malaria is a disease which is pretty much contracted in Indonesia. This is because Indonesia is a tropical region. Malaria is wide spread in tropical and sub tropical regions including part of the American, Asia and Africa [9]. Malaria is an infectious and parasitic diseases plague that affects many people in tropical and subtropical regions, especially in swampy areas. The infected of malaria often peaks during the rainy season [6].

Malaria parasite are transmited by female anopheles mosquitoes bite. The types of protozoa that found in malaria arethe types of *Plasmodium* that are nested within the red blood cells. The parasites multiply within red blood cell causing symptoms that inlude anemia as well as other general symptoms such as fever, chills, flu and in severe cases coma and death. Treatment of malaria are resistantlately. The presence of resistant led to increased the mortality rates. There have reports of treatment resistance of malaria(chloroquine. sulfadoxine-pyrimethamine and quinine) that occurring in over 25% of provinces in Indonesia. Prevention of the development of this resistance can be done with a combination of two or more antimalarial drugs.

WHO (2000), have suggested that Artemesine Based Combination (ACT) as a drug in the treatment of malaria. Artemisin is a sesquiterpene lactone that isolated from the plant

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Artemisia annua. Combination antimalarial therapy has good the activity up to 100% during the 3 days of treatment.

Antimalarials such as artemisinin combination with extracts of natural materials are still rare, but there are a lot of natural materials that a source of antimalarial. Taking preventif drugs (orthodox and traditional drugs) can reduce risk of infection [4]. Antimalarial combination therapy with artemisin also been carried out by [2] that there is a malarial activity of the combination therapy of artesunate and 80% ethanol extract of the stem bark Cempedak (*Artocarpus Champeden* Spreng.). In these studies obtained that the highest percent inhibition of parasite from the extract of *Artocarpus Champeden* is combination extract100 mg/KgBW dan artesunat 36,4 mg/KgBW.

Mudi and bukar (2011), found that there antimalarial activity of the leaf extract *Calotropis* procera. Calatropis gigantea as a genus of plants with Calatropis procera also indicated as having antimalarial activity. This study aims to determine the effect of synergistic combination of 80% ethanol extract of Calatropis gigantea radix and artemisin as antimalarials and to determine the active compounds that contained in the 80% ethanol extract of Calatropis gigantea radix based of reagent test and reinforced by Thin Layer Chromatography (TLC) test.

MATERIALS AND METHODS

Sample Preparation

Radixs of *Calotropis gigantea* were collected at Pasuruan town, Indonesia on January 6, 2014. The radixs were identified at the Departement of Biological Science, University of Malik Ibrahim Malang. The radixs were cleaned and drined. The dried sample was pulverized until 60 mesh.

Extraction of Maseration

The air-dried sample (100 gr) was extracted using Ethanol 80% (250 mL) until 24 hours and filtrated. The residue obtained after extraction was reextracted with Ethanol 80% (150 and 100 mL) or until the filtrat was transparent. The extract were drained and concentrated under reduce pressure using Rotary Evaporator (70-75°C). The ethanol extract was allowed to dry and its contstant weight was recorded.

Chemical Analysis

Plant extract were phytochemically screened using standard techniques for the qualitative detection of Flavonoid, Tannins, Alkaloid, Triterpenoid, Steroid and Saponins. [3] and with TLC (Thin Layer Chromatography) methode.

Antimalarial Activity In Vivo of the Extract on Plasmodium berghei

Antimalarial activity *in vivo* was tested to animal model. Mice were peritoneal infected with 10⁶*Plasmodium berghei* ANKA and divided into 5 treatment groups: negative controll (CMC Na 1%); positive control (artemisin of dose 4 mg/kgBW); combination of artemisin(1/4 dose of artemisin standar) and *Calotropis gigantea*in 3 doses: 0,1 mg/KgBW; 1 mg/KgBWand 10 mg/KgBW. Each group has 6 mice. The parasitemia degree was determined by tail's blood smear of mice until 4 days.

Analysis of Data

The data was described with table and graph. The dose effectivity of 50% (ED50) was determined by percent of probit analyze from parasite inhibition during 4 days and that showed by linear regression analyze with Microsoft Office Excel program. The data of

inhibition was analyzed using SPSS program with *Two Way* ANOVA Test dan continued with Tukey Test.

RESULT AND DISCUSSION

Sample Preparation

Drying sample were meant to reduce the water content, stop the enzymatic reaction, and prevent the growth of mold, so it can be kept longer (preservation) and the chemical composition does not change easily. The dried sample was pulverized until 60 mesh. Pulverizing can facilitate the extraction process. The smaller of surface area are make the greater interaction between extract and solvent, so the extraction process is more effective. The sample with high smoothing powder is also have high probability of damage cells, so it make easier to capture the content material by the solvent.

Extraction of Maseration

The powder samples were weighed as much as 50 g with 2 replications, and its was extracted by ethanol 80% until 24 hours. The maseration process was supported by shaker until 3 hours (120 rpm). The stirring of maceration process is intended to improve the efficiency of solvent saturation, so it will happen faster and more homogeneous extract obtained. In this proces, the solvent will break through the cell of sample, so the active components will be dissolved. The characteristic of active components is same as the characteristic of solvent. It was appropriate by the law of "like dissolve like" that is the only polar compounds that are soluble in polar solvents and non-polar compound is only soluble in non-polar solvents.

The extract were drained and concentrated under reduce pressure using Rotary Evaporator (70-75°C; ±800 hPa; 5 rpm).

Table 1. Results of extraction maceration (*Calotropis gigantea*)

Solvent	Discoloration filtrate	concentrated extract	% (b/b)
Ethanol	Dark green into pale green	Hijau tua	16,4718

Chemical Analysis

Phytochemicals Test with Reagent

Phytochemical test was conducted to determine the content of the ethanol extract compounds in radix of *Calotropis gigantea*. The test was done by taking a sample of ethanol extract. Phytochemical test carried out on this class of compounds Flavonoids, Tannins, Alkaloids, Triterpenoids, Steroids and Saponins.

Table 2. Test 80% ethanol extract of phytochemicals (Calotropis gigantea)

Golongan senyawa	Ekstrak etanol 80%
Flavonoids	-
Tannins	++
Alkaloids	-
Triterpenoids	-
Steroids	++
Saponins	-

Phytochemicals Test with Thin Layer Chromatography (TLC)

The result of phytochemicals test with reagent are tannis and steroids. The verification content of these compounds reinforced with the identification using TLC. The resulting stain subsequently detected with appropriate reagents class compounds, then observed under UV light.

Steroids

The results of identification using TLC for group of steroid compounds in *Calotropis gigantea* using eluent hexane: acetone (3.5: 1.5) that showed by Figure 1. Reveny (2011), shows that the results of the TLC group of steroid compounds with Lieberman-Burchard reagent showed by the formation of a red purple. In the ethanol extract showed Rf between 0,05 to 0,6 with 7 stain. The 1^{st} and 2^{nd} stain showed a Greenish white, and the $3^{rd} - 7^{th}$ stain showed a red purple.

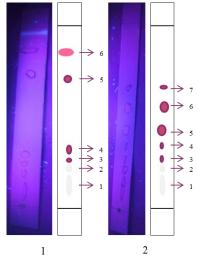


Fig 1. The results of the ethanol extract of steroid compounds TLC eluent hexane: acetone (3.5: 1.5), (1) Results spotting before sprayed detection reagents (366 UV lamp), (2) Results spotting after sprayed detection reagents (366 UV lamp)

Table 3. The results of steroid TLC with eluent hexane: acetone (3.5: 1.5)

No	Rf	Color patches with	Rf	Color patches with 366	Alleged
		366 nm UV light,		nm UV light, after given	compound
		before given detector		detector	
1	0,05	Grayish-white	0,05	Greenishwhite	-
2	0,16	Grayish-white	0,16	Greenish white	-
3	0,22	Red purple	0,22	Red purple	Steroid
4	0,26	Red purple	0,26	Red purple	Steroid
5	0,76	Red purple	0,33	Red purple	Steroid
6	0,86	Pink	0,38	Red purple	Steroid
7			0,49	Red purple	Steroid
8			0,6	Red purple	Steroid

Tannins

The results of identification using TLC for group of tannins compounds in *Calotropis gigantea* using eluent chloroform: methanol: water (7:3:0,4)that showed by Figure 2. Fitriyani *et al* (2011), shows that the results of the TLC group of tannins compounds with

FeCl₃ reagent showed by the formation of a blackish purple.

Table 4. The results of tannins TLC with eluent chloroform: methanol: water (7:3:0,4)

No	Rf	Color patches with 366	Rf	Color patches with	Alleged
		nm UV light, before		366 nm UV light, after	compound
		given detector		given detector	
1	0,62	Red Purple	0,41	greenish white	-
2	0,78	Grayish-white	0,62	Blackish purple	Tanin
3	0,85	Blackish purple	0,78	Grayish-white	-
4			0,85	Red Purple	-
5			0,87	Cream	-

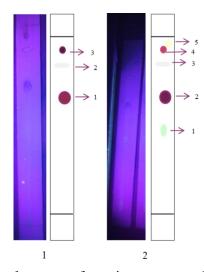


Fig 2. The results of the ethanol extract of tannins compounds TLC chloroform: methanol: water (7:3:0,4), (1) Results spotting before sprayed detection reagents (366 UV lamp), (2) Results spotting after sprayed detection reagents (366 UV lamp)

Antimalarial Activity In Vivo of the Extract on Plasmodium berghei

The test of antimalarial activity in vivo performed withanimal tests using male mice (*Mus musculus*; Balb/C). Mice were peritoneal infected with 10⁶*Plasmodium berghei* ANKA and divided into 5 treatment groups: negative control; positive control (artemisin of dose 4 mg/kgBW); combination of artemisin(1/4 dose of artemisin standar) and *Calotropis gigantea*in 3 doses: 0,1 mg/KgBW; 1 mg/KgBWand 10 mg/KgBW.

Table 5. Parasitemia average degree of radix extract (*Calotropis gigantea*)

Treatment	Average degree of parasitemia(%) ± standard deviation				
	Day-0	Day-1	Day-2	Day-3	Day-4
Control (-)	$2,9\pm0,76$	$3,04\pm0,22$	$3,4\pm0,33$	$3,6\pm1,04$	$3,86\pm0,33$
Control (+)	$2,34\pm0,43$	$2,22\pm0,43$	$1,54\pm0,30$	$1,38\pm0,22$	$0,7\pm0,11$
Dose 1	$3,88\pm0,48$	$2,34\pm0,33$	$1,7\pm0,50$	$1,44\pm0,35$	$0,5\pm2,86$
Dose 2	$3,14\pm0,60$	$2,12\pm0,85$	$1,74\pm0,31$	$1,9\pm0,51$	$1,04\pm0,43$
Dose 3	$2,4\pm1,26$	$2,32\pm0,39$	$1,74\pm0,29$	$1,2\pm0,10$	$1,72\pm0,23$

Therapy was done when the degree of parasitemia after infection reaches 1-5%(it calculated as day 0). The degree of parasitemia on day 0 is aiming to prove all the mice are in the same range of degrees of parasitemia on day of treatment will be done. Therapy was given for 4 days, that use to know the profile of parasite growth after a given treatment.

The average degree of parasitaemia in all treatments on day 0 is equal to 2-3%. Treatment of the negative control group showed the highest degree of parasitemia. The degree of parasitemia was obtained from a blood thinner dosage by counting the number of cells infected with *Plasmodium berghei* in 1000 erythrocytes. The observation of blood smear Plasmodium berghei infected in the negative control (no treatment) are presented in Figure 3.

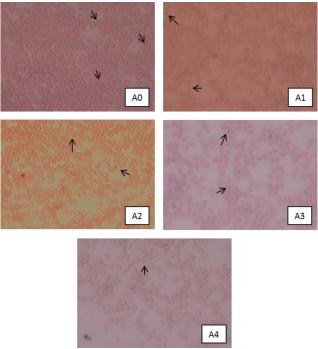


Fig 3. Picture of erythrocytes infected control (-) / no treatment on day 0, day 1, day 2, day 3, and day 4, with a 400x magnification microscope observations.

The number of erythrocytes was infected by Plasmodium berghei increased along with increasing the treatment. The cells that infected with Plasmodium berghei will form a ring-shaped trophozoites. The erythrocytes that containing schizonts and tropozoit are have rough spots (indicated by arrows).

The treatment process can reduce the degree of parasitemia. Degree of parasitemia is used to determine the percent inhibition that resulting from any dose. The dose of effectiveness can be seen from the% inhibition in each individual treatment group. % Inhibition can be determined by the formula.

% inhibition=
$$\frac{(parasitemia\ of\ negative\ control\ -\ parasitemia\ of\ extract)}{parasitemia\ of\ negative\ control} x\ 100\ \%$$

Treatment	% Inhibition	Inhibition probit
Dose 0,1 mg/Kg BB	55,22	5,15
Dose 1 mg/Kg BB	72,866	5,74
Dose 10 mg/Kg BB	87,342	6,182

Table 6. The average value of percent and probit inhibition (Day-4)

The dose effectivity of 50% (ED50) was determined by percent of probit analyze from parasite inhibition during 4 days and that showed by linear regression analyze with Microsoft Office Excel program. The dose effectivity of 50% (ED50) can not be determined because the average percent inhibition was above 50% (55% - 87%). Pouplin *et al.* (2007), an extract is said to have the nature antiplasmodium when parasite inhibition may provide more than 30%. It can be concluded that the combination of *Calotropis gigantea* radix and artemisin have a very good potential in inhibiting the growth of the parasite.

Inhibition of *Plasmodium berghei* was due to the active compounds that can be contained from ethanol extract of *Calotropis gigantea* radix. The active compounds that can inhibit the growth of the parasite are steroids and tannins.

CONCLUSION

Conclusively, the combination of *Calotropis gigantea* radix and artemisin has a synergistic effect as an antimalarial drug. The result shows that combination of *Calotropis gigantea* radix extract and artemisin with dose 10 mg/KgBW has higher antimalarial activity than control positive (artemisin). The value of parasite inhibition is 55,2% for dose 0.1 mg/KgBW; 72,8% for dose 1 mg/KgBW; 87,3% for dose 10 mg/KgBW and 56,06 mg/KgBW for control positive (artemisin).

The active compounds that contained from ethanol extract of *Calotropis gigantea* radix with reagent test are steroids and tannins. It is also supported by the results using the TLC separation, the separation of steroid compounds that produce a reddish purple color with Rf 0.22; 0.26; 0.33; 0.38; 0.49 and 0.6 and tannin produces purple-black color with Rf 0.62.

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REFERENCES

- [1] Fitriyani, A, dkk. 2011, Jurnal Majalah Obat Tradisional, 16(1), 34-42.
- [2] Hafid F.A., Tyas W.M dan Widyawaruyanti A. 2011, J Indon Med Assoc, Vol.61. No 4.
- [3] Kristanti A.N, Aminah N.S, Tanjung M, dan Kurniadi B. 2008. *Buku Ajar Fitokimia*. Surabaya: Airlangga University Pers. HX Chen; J Ying; H Chen; JL Huang; L Liao, Chromatographia, 2008, 6, 629-634.
- [4] Mary J.M, Richard A.H, Pamela C.C, Bruce D.F, Michael C. 2000. Lippincott's Illustrated Reviews: *Pharmacology*. Lippincott Williams & Wilkins, A Walter Kluwer Company, Philadelphia, USA, 349-353.
- [5] Mudi, Y.S dan Bukar. A. 2011, *Nigerian Society for Experimental Biology*, Vol. 22. No. 1.P Vinas; N Balsalobre; MH Cordoba, Analytical Chimica Acta, 2006, 558, 11–15.

- [6] Newman, J.L., Mahretu, A., Shillington, K., and Stock, R. 2008. *Africa*. Microsoft Student 2008 [DVD]. Redmond, WA: Microsoft Coorperation.
- [7] Pouplin, J.N., T.H., Tran, C., Dolecek, T.A., Phan, J., Farrar, P., Carron, B., Bodo, & P., Grellier, 2007, *Journal of ethnopharmacology*, Vol. 109.A Gantverg; I Shishani; M Hoffman, Analytica Chimica Acta, 2003, 483, 125-135.
- [8] Reveny, Julia (2011), Jurnal ILMU DASAR, Vol. 12 No. 1, Januari 2011: 6-12.
- [9] Snow R.W, Guerra CA, Noor Am, Myint Hy, Hay SI. 2005, Nature 434 (7030): 214-7.
- [10] WHO. 2010. Guidlines for The Treatment of Malaria. WHO Geneva.